PII: S0957-4166(97)00372-8

Microbial reduction of 2-(6-m-methoxyphenyl-3-oxohexyl)-2,4,4-trimethylcyclopenta-1,3-dione with *Schizosaccharomyces pombe* (NRRL Y-164)

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Abstract: Racemic 2-(6-m-methoxyphenyl-3-oxohexyl)-2,4,4-trimethylcyclopenta-1,3-dione 4 was resolved by reduction with Schizosaccharomyces pombe (NRRL Y-164) to give (-)-5, (+)-6, and (+)-7 in 37, 12, and 19% yields, respectively. Chromic acid oxidation of (-)-5 gave (+)-4 while oxidation of (+)-6 and (+)-7 gave (-)-4, respectively. Sodium borohydride reduction of (+)-4 followed by Oppenauer oxidation yielded (-)-5 and (-)-7, and the same treatment of (-)-4 yielded (+)-5 and (+)-7, respectively. © 1997 Elsevier Science Ltd

Introduction

In a previous communication, 1 we reported the resolution of racemic 2-(6-m-methoxyphenyl-3-oxohexyl)-2,4,5-trimethylcyclopenta-1,3-dione 1 by microbial reduction with Schizosaccharomyces pombe (NRRL Y-164) to give 2 and 3 in 42 and 36% yields, respectively (Scheme 1). Racemic 2-(6-m-methoxyphenyl-3-oxohexyl)-2,4,4-trimethylcyclopenta-1,3-dione 4 was prepared earlier by us and subjected to an asymmetric cyclization reaction with L(-) and D(+)-phenylalanine as the asymmetry inducers, 2 respectively. In view of the successful resolution of 1 with S. pombe (NRRL Y-164), 4 was incubated with this same microorganism. We would like to report the results obtained together with the chemical correlation reactions performed during the characterization of transformation products.

* Schizosaccharomyces pombe (NRRL Y-164)

Scheme 1.

Results and discussion

From the incubation mixture of (\pm) -4 with S. pombe (NRRL Y-164), (-)-5, (+)-6, and (+)-7 were isolated in 37, 12, and 19% yields, respectively (Scheme 2).

Compound (-)-5, colorless oil with $[\alpha]_D^{25}$ -19.0 (c 0.94, CHCl₃), has a molecular formula $C_{21}H_{30}O_4$ as deduced from its HREIMS m/z 346.2146 (calcd 346.2144). Its IR absorption at 3500 cm⁻¹ and a carbinoyl proton signal at δ 4.04 (dd, J=6.5 and 8.9 Hz) in its ¹H-NMR spectrum support the presence of a secondary hydroxyl group on the five-membered ring. The fact that (-)-5

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(I) Schizosaccharomyces pombe (NRRL Y-164)

(II) CrO₃-H⁺; (III) Ac₂O-Py

Scheme 2.

is resistant to Oppenauer oxidation³ but can be oxidized by Jones' reagent⁴ to (+)-4, $[\alpha]_D^{24}$ +4.9 (c 1.0, CHCl₃), support this suggestion. Therefore, 1-hydroxy-2-(6-*m*-methoxyphenyl-3-oxohexyl)-2,4,4-trimethylcyclopenta-3-one was assigned to (-)-5 and its (1*S*,2*R*)-stereochemistry was determined by chemical correlation which will be described later.

Compound (-)-6, $[\alpha]_D^{25}$ +83 (c 1.0, CHCl₃), mp 95-96°C, has a molecular formula $C_{21}H_{30}O_4$ as deduced from its HREIMS m/z 346.2141 (calcd 346.2144). Its IR absorption at 3420 and 1730 cm⁻¹ suggested the presence of a hydroxyl and a carbonyl function. The ¹³C-NMR spectrum revealed a sole carbonyl carbon (δ 223.3), a dioxygenated carbon (δ 102.2) and an oxygenated methine (δ 69.8, d) which couples directly to a carbinoyl proton (δ 3.70), identified by a HMOC spectrum. The molecular formula of (+)-6 provided seven ring and double bond equivalents, six of which are easily identified from the presence of an aryl ring, a carbonyl group and the five-membered ring in the molecule. The HMBC spectrum (Figure 1) showed that the dioxygenated carbon coupled to 6-Me singlet (δ 1.01) and 9-H₂ (δ 1.94 and 2.10), and the carbonyl carbon coupled to 6-Me, 8 α - and 8 β -Me. This afforded the exact substitution pattern for the five-membered ring and the dioxygenated carbon to be a hemiketal carbon, probably ether-linked to C-3 to form a pyran ring accounting for the seventh equivalent. This suggestion was supported by the coupling pattern of the carbinoyl carbon (H-3, δ 3.70, ddt, J=11.4, 1.5, 5.6 Hz) which indicated H-3 to be an axial proton coupling to the flexible C-1' proton (δ 1.40), and the axial proton (δ 0.96, J=11.4 Hz) and equatorial proton (δ 1.42, m) of C-4, clarified by a COSY-45 spectrum. Based on these analyses and supported by the X-ray diffraction analysis⁵ the structure of (+)-6 was established as (15.3R.6R)-1-hydroxy-3-(3-m-methoxyphenylpropyl)-6,8,8trimethyl-2-oxabicyclo[4,3,0]nonan-7-one.

Compound (+)-7, $[\alpha]_D^{25}$ +18.8 (c 1.0, CHCl₃), has a molecular formula $C_{21}H_{30}O_4$ as deduced from its HREIMS m/z 346.2144 (calcd 346.2144). Its IR absorption at 3450 and 1740 cm⁻¹ suggested the presence of a hydroxyl and a carbonyl function as similar to (+)-6. The ¹³C-NMR spectrum revealed a sole carbonyl carbon (δ 225.5), a dioxygenated carbon (δ 96.4, s) and an oxygenated methine (δ

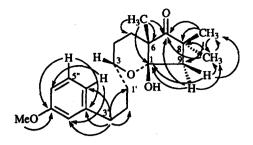


Figure 1. HMBC correlations for (+)-6.

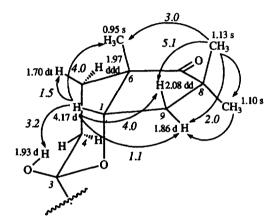


Figure 2. Partial ¹H-NMR data and NOED (arrow, %) of (+)-7.

75.5, d) which couples directly to a carbinoyl proton (δ 4.17), identified by an HMQC spectrum. These data suggested that (+)-7 is structually related to (+)-6. The ¹H-NMR spectrum showed an D₂O exchangeable doublet at δ 1.93 (J=2.0 Hz), which couples to H-4 α (an axial proton) (δ 1.22, ddt, J=2.0, 4.18, 13.4 Hz), identified by the D₂O exchange experiment and double resonance at δ 1.93, both of which collapsed H-4 α to a double triplet (J=4.8, 13.4 Hz). These results locate 3 β -OH which H-bonds to pyran oxygen (O-2) to make a W-shape coupling between OH and H-4 α . An NOE study enhancing 3 β -OH, 6-Me (δ 0.95, s), H-9 α (δ 1.86, d) upon irradiation of H-1 (δ 4.17) also confirmed the *cis* relationship of H-1 to both 3 β -OH and 6-Me (Figure 2). Treatment of (+)-7 with Ac₂O-Py (r.t., 18h) yielded a needle shaped crystalline (+)-8, [α]_D²⁵ +6.8 (c 1.25, CHCl₃), mp 58–59°C, having a molecular formula C₂₃H₃₂O₅ as deduced from its HREIMS m/z 388.2242 (calcd 388.2250). The structure of (+)-8 was elucidated as (+)-(1S,2S)-1-acetoxy-2-(6-m-methoxyphenyl-3-oxohexyl)-2,4,4-trimethylcyclopenta-3-one based on X-ray diffraction analysis. Based on the data obtained in (+)-8, the structure of (+)-7 was concluded to be (1S,3R,6S)-3-hydroxy-3-(3-m-methoxyphenylpropyl)-6,8,8-trimethyl-2-oxabicyclo[4,3,0]nonan-7-one. Detailed ¹H- and ¹³C-NMR assignments of (+)-7 were made by analysis of COSY-45, HMQC, and HMBC spectra (Figure 3).

Chromic acid oxidation of both (+)-6 and (+)-7 yielded (-)-4, $[\alpha]_D^{25}$ -5.0 (c 1.0, CHCl₃), respectively. From earlier experience,² the carbonyl group flanked by four vicinal alkyl groups on the cyclopentane ring was not reduced under the condition of NaBH₄/MeOH/0°C.⁷ This is a similar result to that which we obtained here with *S. pombe* (NRRL Y-164) and (±)-4. The NaBH₄ reduction of (+)-4 followed by Oppenauer oxidation³ yielded two mono-alcohols in 28 and 42% yields, respectively. The minor product with $[\alpha]_D^{25}$ -19.2 (c 1.04, CHCl₃) is identical in every respect (UV, IR and NMR) to (-)-5 isolated from microbial reduction. The major product with $[\alpha]_D^{25}$ -18.9 (c 1.06, CHCl₃),

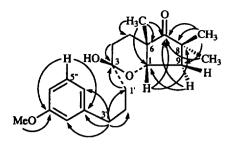


Figure 3. Major HMBC correlation for (+)-7.

possessed identical spectral data to (+)-7 but with opposite optical rotation. The same treatment of (-)-4 also yielded a minor product (+)-5 (28%) with $[\alpha]_D^{25}$ +19.1(c 1.02, CHCl₃), and a major product (+)-7 (43%) with $[\alpha]_D^{25}$ +18.7 (c 0.99, CHCl₃) (Scheme 3). During these chemical correlation reactions with (+)- and (-)-4 as the starting material, respectively, a pair of epimeric cyclopentanols formed in each case. In both instances, the major product was the one with the hydroxy group *trans* oriented in regard to the C-2 methyl which formed a pyrane ring involving the C-3' side-chain carbonyl and leading to the formation of a hemiketal carbon. The minor product was the one with the hydroxy group *cis* oriented in regard to the C-2 methyl and remained unchanged. Based on results obtained on chemical correlation, the (15,2R)-stereochemistry for (-)-5 was assigned. These results also indicated the essential optical purity of these microbially reduced products.

(+) -4
$$\xrightarrow{(i), (ii)}$$
 (-) -5 (28%) + H₃CO \xrightarrow{i} H (-) -7 (42%) [α]_D²⁵ = -18.9

(-)-4
$$\xrightarrow{(i), (ii)}$$
 + $(+)-7 (43\%)$ | $[\alpha]_D^{25} = +18.7$ | $[\alpha]_D^{25} = +19.1$

(i) NaBH₄, MeOH, 0°C , 2h. (ii) Al(i-PrO)₃, Me₂CO-C₆H₆, reflux, 21h.

Scheme 3.

Thus, in this study, we showed that resolution of (\pm) -4. could be achieved by microbial reduction with S. pombe (NRRL Y-164) in moderate yields. In both enantiomers, the less sterically hindered carbonyl group on the cyclopentane ring could be reduced to (S)-alcohol but to a different extent. In (+)-4, the less hindered carbonyl group is located in such a way that it is equivalent to the C-17 in steroid and was reduced more extensively. On the other hand, in (-)-4, the side-chain carbonyl was also reduced to a considerable extent.

Experimental

General

Melting points measured on a Buchi 510 melting point apparatus were uncorrected. Optical rotations were measured on a Jasco DIP-370 digital polarimeter. UV spectra were recorded on a Hitachi 150-20 double beam spectrometer. IR spectra were recorded on a Jasco A-100 infrared spectrophotometer. 1 H- and 13 C-NMR spectra were obtained on Bruker AM-200 or Bruker AMX-400 spectrophotometer in CDCl₃ or CD₃OD using the solvent peaks as reference standards. 2D-NMR spectra were recorded by using Bruker's standard pulse program: in the HMQC and HMBC experiments, Δ =1 s and J=140, 8 Hz, respectively, the correlation maps consisted of 512×1 K data points per spectrum, each composed of 16 to 64 transients. EIMS were recorded on Finnigan Mat 4500 GC/MS and HREIMS on JEOL JMX-HX-110 mass spectrometer at 70 eV. Normal phase silica gel for column chromatography were purchased from Merck, 7734 (70–230 mesh) and 9385 (230–400 mesh); TLC plates of Merck 573 (Si60 with F_{254} , 0.25 mm) were purchased from E. Merck, A.G., Darmstadt, Germany.

All of the solvents and inorganic chemicals were reagent grade. Sodium borohydride and aluminum isopropoxide were purchased from Aldrich chemical Co., Inc., Milwaukee, WI. Nutrient broth, maltose, proteose peptone *3 and bacto-agar were from Difco Laboratories, Detroit, MI. Dextrose was obtained from Wako Pure Chemical Industries, Ltd, Osaka, Japan.

The microorganism, Schizosaccharomyces pombe (NRRL Y-164), was maintained on an MP-#3 Agar (Maltose 4%, Proteose Peptone #3 1.5%, and Agar 3%) at 26°C for 11 days then transferred and grown in Nutrient broth-Dextrose medium (Nutrient broth 1.6% and Dextrose 4%) at 24–26°C on a rotary shaker (250 rpm, 1 in. stroke). Transformations were carried out in 2 L Erlenmyer flasks containing 400 mL of the same medium.

Microbiological reduction of (\pm) -4

The microorganism was grown in 8 L of Nutrient broth–Dextrose medium contained in twenty 2 L Erlenmyer flasks. (±)-4 (1.83 g) dissolved in 40 mL DMF was added and the incubation was continued for 78 h. Extraction of the incubation mixture with CHCl₃ (3×2.7 L) and subsequent work-up gave a yellow oily residue (2.8 g). The residue was chromatographed over a silica gel column (400 g) and eluted with CHCl₃-Me₂CO mixture. The fractions (0.3 g) obtained from CHCl₃ elution were further purified over another silica gel column to recover unreacted substrate, (±)-4 (240 mg, 13%). The fractions (0.25 g) obtained from CHCl₃-Me₂CO (99:1) were further purified over another silica gel column to give pure (+)-6 (220 mg, 12%). The fractions (0.4 g) obtained from CHCl₃-Me₂CO (97:3) were further purified over another silica gel column to give pure (+)-7 (349 mg, 19%). The fractions (0.7 g) obtained from CHCl₃-Me₂CO (95:5) were further purified over another silica gel column to give pure (-)-5 (679 mg, 37%).

(1S,2R)-1-Hydroxy-2-(6-m-methoxyphenyl-3-oxohexyl)-2,4,4-trimethyl-cyclopenta-3-one (-)-5

Colorless liquid; $[\alpha]_D^{25}$ – 19.0 (c=0.94, CHCl₃); IR (CHCl₃) ν_{max} 3500 (OH), 2960, 1735, 1715 (C=O), 1600, 1585, 1490, 1460, 1370, 1260, 1155 and 1045 cm⁻¹; UV (MeOH) λ_{max} (log ε) 272 (3.26), 279 (3.22) nm; ¹H-NMR δ (CDCl₃) 7.15 (1H, t, *J*=7.8 Hz, H-5"), 6.71 (1H, br d, *J*=7.8 Hz, H-6"), 6.70 (1H, br d, *J*=7.8 Hz, H-4"), 6.68 (1H, br s, H-2"), 4.04 (1H, dd, *J*=8.9, 6.5 Hz, H-1), 3.75 (3H, s, 3"-OMe), 2.54 (2H, t, *J*=7.5 Hz, 6'-H), 2.38 (4 H, t, *J*=7.2 Hz, H-2" and H-4"), 2.05 (1H, dd, *J*=12.8, 6.5 Hz, H-5α), 1.85 (2H, quintet, *J*=7.5 Hz, H-5'), 1.76 (1H, dd, *J*=12.8, 8.9 Hz, H-5β), 1.74 (2H, m, H-1"), 1.12 (3H, s, 4β-Me), 0.94 (3H, s, 4α-Me), 0.93 (3H, s, 2-Me); ¹³C-NMR δ (CDCl₃) 224.0 (s, C-3), 211.2 (s, C-3'), 159.6 (s, C-3"), 143.0 (s, C-1"), 129.3 (d, C-5"), 120.8 (d, C-6"), 114.2 (d, C-2"), 111.2 (d, C-4"), 73.0 (d, C-1), 55.0 (q, 3"-OMe), 52.8 (s, C-2), 44.6 (s, C-4), 42.7 (t, C-5), 41.8 (t, C-4'), 37.6 (t, C-2'), 35.0 (t, C-6'), 28.6 (t, C-1'), 24.9 (t, C-5'), 26.0 (q) and 25.2 (q) (4α- and 4β-Me), 15.8 (q, 2-Me); NOED (CDCl₃) 2-Me to H-1" (17.1%), 4α-Me to H-1 (6.8%), H-5α (5.6%) and 4β-Me (7.3%), 4β-Me to H-5β (3.6%) and 4α-Me (9.1%); HREIMS m/z

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346.2146 (calcd for $C_{21}H_{30}O_4$ 346.2144); EIMS m/z [M]⁺ 346 (20), 328 (20), 212 (22), 197 (29), 177(25), 141 (50), 134 (100), 121 (18).

(1S,2R,6R)-1-Hydroxy-3-(3-m-methoxyphenylpropyl)-6,8,8-trimethyl-2-oxabicyclo[4,3,0]nonan-7-one (+)-6

Colorless needle crystals, mp 95–96°C; $[\alpha]_D^{25}$ +83.0 (c=1.0, CHCl₃); IR (KBr) ν_{max} 3420 (OH), 2950, 1730 (C=O), 1610, 1585, 1490, 1455, 1440, 1365, 1305, 1260, 1160, 1105 cm⁻¹; UV (MeOH) λ_{max} (log ϵ) 273 (3.36), 279 (3.32) nm; ¹H-NMR δ (CDCl₃) 7.16 (1H, dd, J=8.9, 7.6 Hz, H-5"), 6.74 (1H, br d, J=7.6 Hz, H-6"), 6.70 (1H, br d, J=7.6Hz, H-4"), 6.69 (1H, br s, H-2"), 3.77 (3H, s, 3"-OMe), 3.70 (1H, m, H-3), 2.55 (2H, t, J=7.7 Hz, H-3'), 2.10 (1H, d, J=12.8 Hz, H-9 β), 2.02 (1H, ddd, J=13.6, 4.0, 2.8 Hz, H-5 α), 1.94 (1H, d, J=12.8 Hz, H-9 α), 1.42 (1H, m) and 0.96 (1H, m) (H-4), 2.00 (1H, m) and 1.50 (1H, m) (H-2'), 1.40 (2H, m, H-1'), 1.17 (3H, s, 8 β -Me), 1.13 (3H, s, 8 α -Me), 1.01 (3H, s, 6-Me); ¹³C-NMR δ (CDCl₃) 223.3 (s, C-7), 159.7 (s, C-3"), 144.1 (s, C-1"), 129.2 (d, C-5"), 120.8 (d, C-6"), 114.3 (d, C-2"), 110.9 (d, C-4"), 102.2 (s, C-1), 69.8 (d, C-3), 55.1 (q, 3"-OMe), 51.8 (s, C-6), 48.7 (t, C-9) and 45.0 (s, C-8), 35.9 (t, C-3'), 35.5 (t, C-1'), 28.5 (t, C-4), 28.0 (q, 8 α -Me), 27.7 (t, C-5), 26.9 (t, C-2'), 26.5 (q, 8 β -Me), 21.8 (q, 6-Me); NOED (CDCl₃) 6-Me to H-5 α (4.4%), H-5 β (6.4%), 8 β -Me (2.9%) and H-9 β (5.8%), 8 α -Me to H-9 α (5.9%) and 8 β -Me (5.0%), 8 β -Me to 6-Me (2.6%), H-9 β (7.0%) and 8 α -Me (6.5%); HREIMS m/z [M]* 346.2141 (calcd for C₂₁H₃₀O₄ 346.2144); EIMS m/z [M]* 346 (15), 328 (12), 161 (30), 141 (20), 134 (100), 121 (30), 109 (8).

(1S,3R,6S)-3-Hydroxy-3-(3-m-methoxyphenylpropyl)-6,8,8-trimethyl-2-oxabicyclo[4,3,0]nonane-7-one (+)-7

Colorless liquid; $[\alpha]_D^{25}$ +18.8 (c=1.0, CHCl₃); IR (CHCl₃) ν_{max} 3450 (OH), 2950, 1740 (C=O), 1600, 1580, 1495, 1460, 1380, 1260, 1155, 1060 cm⁻¹; UV (MeOH) λ_{max} (log ε) 272 (3.26), 279 (3.21) nm; ¹H-NMR δ (CDCl₃) 7.16 (1H, t, *J*=7.8 Hz, H-5"), 6.72 (1H, br d, *J*=7.8 Hz, H-6"), 6.70 (1H, br d, *J*=7.8 Hz, H-4"), 6.68 (1H, br s, H-2"), 4.17 (1H, d, *J*=4.4 Hz, H-1), 3.76 (3H, s, 3"-OMe), 2.54 (2H, t, *J*=7.7 Hz, H-3'), 2.08 (1 H, dd, *J*=14.1, 4.4 Hz, H-9β), 1.97 (1H, ddd, *J*=13.4, 4.8, 2.8 Hz, H-5α), 1.93 (1H, d, *J*=2.0 Hz, 3-OH), 1.86 (1H, d, *J*=14.1 Hz, H-9α), 1.70 (1H, dt, *J*=4.6, 13.4 Hz, H-5β), 1.66 (2H, m, H-2'), 1.55 (2H, m, H-1'), 1.51 (1H, ddd, *J*=13.4, 4.6, 2.8 Hz, H-4β), 1.22 (1 H, ddt, *J*=2.0, 4.8, 13.4 Hz, H-4α), 1.13 (3H, s, 8β-Me), 1.10 (3H, s, 8α-Me), 0.95 (3H, s, 6-Me); ¹³C-NMR δ (CDCl₃) 225.5 (s, C-7), 159.6 (s, C-3"), 143.6 (s, C-1"), 129.3 (d, C-5"), 120.8 (d, C-6"), 114.2 (d, C-2"), 111.1 (d, C-4"), 96.4 (s, C-3), 75.5 (d, C-1), 55.1 (q, 3"-OMe), 49.4 (s, C-6), 43.4 (s, C-8), 42.6 (t, C-1'), 41.1 (t, C-9), 35.8 (t, C-3'), 30.0 (t, C-4), 28.4 (q, 8β-Me), 27.1 (q, 8α-Me), 24.9 (t, C-5), 24.7 (t, C-2'), 22.5 (q, 6-Me); Major COSY 45: H_α-4+3-OH, H_β-4, H_α-5, H_β-5; H_β-4+H_α-4, H_α-5, H_β-5; H-1+H_β-9+H_α-9; H-1'+H-2'+H-3'; H-4"+H-5"+H-6"; HREIMS m/z [M]⁺ 346.2144 (calcd for C₂₁H₃₀O₄ 346.2144); EIMS m/z [M]⁺ 346 (8), 328 (4), 212 (10), 194 (8), 177 (12), 142 (54), 134 (100), 121 (22), 109 (12).

Chromic acid oxidation of carbinols⁴

In a typical Jones oxidation procedure, a solution of Me_2CO (6 mL) containing the carbinol (60 mg, 0.173 mmol) was cooled to 0° – $5^{\circ}C$, then 0.25 mL (0.6 mmol) of CrO_3 – H_2SO_4 solution (prepared by dissolving 26.8 g of CrO_3 in 23 mL of conc. H_2SO_4 and H_2O added to make 100 mL) was added dropwise with stirring and the reaction mixture was stirred at room temperature for 30 min. Then EtOH (0.5 mL) and H_2O (10 mL) were added to terminate the reaction. The mixture was extracted with $CHCl_3$ (10 mL×3), and the combined $CHCl_3$ extract was washed with saturated NaCl solution and dried over Na_2SO_4 . Evaporation of the solvent under reduced pressure and purification of the product residue over silica gel column gave analytical sample.

(a) Oxidation of (-)-5: (-)-5 (100 mg) was oxidized to give (+)-4 (72.6 mg, 73%), $[\alpha]_D^{24}$ +4.9 (c=1.0, CHCl₃).

- (b) Oxidation of (+)-6: (+)-6 (50 mg) was oxidized to give (-)-4 (35.2 mg, 70%), $[\alpha]_D^{25}$ -5.0 (c=1.0, CHCl₃).
- (c) Oxidation of (+)-7: (+)-7 (50 mg) was oxidized to give (-)-4 (35 mg, 70%), $[\alpha]_D^{25}$ -5.0 (c=1.0, CHCl₃).

(1S,2S)-1-Acetoxy-2-(6-m-methoxyphenyl-3-oxohexyl)-2,4,4-trimethyl-cyclopenta-3-one (+)-8

The (+)-7 (91 mg, 0.25 mmol) dissolved in pyridine (5 mL) was treated with Ac₂O (0.25 mL, 2.45 mmol) and left standing at r.t. for 18 h. To the reaction mixture, H₂O (40 mL) was added and the mixture was extracted with CHCl₃ (15 mL×3). The combined CHCl₃ was washed with H₂O, dried over MgSO₄ and the solvent evaporated to dryness to give crystalline residue. Purification of the residue over a small silica gel column and recrystallization from Me₂CO-Pet, ether gave (+)-8 (76 mg, 74%), mp 58–59°C; $[\alpha]_D^{25}$ +6.8 (c=1.25, CHCl₃); IR (CHCl₃) ν_{max} 2970, 2930, 1735 and 1715 (C=O and ester), 1600, 1580, 1455, 1370, 1240, 1150, 1030 cm⁻¹; UV (MeOH) λ_{max} (log ϵ) 273 (3.36), 279 (3.32) nm; 1 H-NMR δ (CDCl₃) 7.17 (1H, t, J=7.8 Hz, H-5"), 6.73 (1H, br d, J=7.8 Hz, H-6"), 6.70 (1H, br d, J=7.8 Hz, H-4''), 6.69 (1H, br s, H-2''), 5.12 (1H, t, J=5.9 Hz, H-1), 3.77 (3H, s, 3''-OMe),2.56 (2H, t, J=7.6 Hz, H-6'), 2.47 (1 H, ddd, J=17.3, 10.6 and 5.5 Hz,) and 2.28 (1 H, ddd, J=17.3, 10.6 and 5.1 Hz,) (H-2'), 2.37 (2H, J=7.4 Hz, H-4'), 2.18 $(1H, dd, J=13.9, 6.0 Hz, H-5\beta)$, 1.90 $(1H, dd, J=13.9, 6.0 Hz, H-5\beta)$ dd, J=13.9, 5.7 Hz, $H-5\alpha$), 1.87 (2H, m, H-5'), 1.82 (1H, ddd, J=14.5, 11.5 and 5.4 Hz) and 1.70 (1H, ddd, J=14.5, 10.6 and 5.2 Hz) (H-1'), 1.11 (3H, s, 4α -Me), 1.07 (3H, s, 4β -Me), 1.03 (3H, s, 2-Me); ¹³C-NMR δ (CDCl₃) 222.7 (s, C-3), 209.9 (s, C-3'), 170.3 (s, 1-OC (=0) Me), 159.6 (s, C-3''), 143.2 (s, C-1"), 129.3 (d, C-5"), 120.8 (d, C-6"), 114.2 (d, C-2"), 111.2 (d, C-4"), 77.7 (d, C-1), 55.1 (q, 3"-OMe), 51.3 (s, C-2), 44.5 (s, C-4), 42.0 (t, C-4'), 40.6 (t, C-5), 37.0 (t, C-2'), 35.1 (t, C-6'), 25.0 (t) and 25.1 (t) (C-1' and C-5'), 26.5 (q, 4 β -Me), 25.9 (q, 4 α -Me), 21.05 (q, Me of OAc), 20.87 (q, 2-Me); NOED (CDCl₃) 2-Me to H-1' (2.8%), H-2' (d 2.28, 3.7%; d 2.47, 5.2%), H-3 (10.6%) and $H-5\beta$ (3.7%), 4α -Me to $H-5\alpha$ (6.4%) and 4β -Me (9.6%), 4β -Me to H-1 (5.1%), $H-5\beta$ (6.7%) and 4α -Me (19.1%); HREIMS m/z [M]⁺ 388.2242 (calcd for $C_{23}H_{32}O_5$ 388.2250); EIMS m/z 389 (7), $[M]^+$ 388 (29), 370 (6), 254 (27), 194 (7), 184 (51), 177 (48), 151 (10), 134 (100), 124 (72), 109 (17).

The NaBH₄ reduction⁷ followed by Oppenauer oxidation³ of (+)- and (-)-4

- (a) Reaction of (+)-4: The starting material, (+)-4 (302 mg, 0.87 mmol), was dissolved in MeOH (10 mL) and cooled to 0°C. NaBH₄ (340 mg, 8.7 mmol) was added to it in portions with stirring, the reaction was terminated after 2 h by addition of H₂O (40 mL) and extracted with CHCl₃ (20 mL×3). The combined CHCl₃ extract was washed with H₂O, dried over Na₂SO₄ and the solvent removed *in vacuo* to dryness. The residue was purified over a silica gel column (20 g) eluted with CHCl₃-Me₂CO mixture to give 300 mg of residue after evaporation of solvent. The residue was dissolved in Me₂CO-C₆H₆ (10:15 mL) and after addition of Al(Me₂CHO)₃ (244 mg, 1.0 mmol) the mixture was heated at reflux for 21 h. After cooling, H₂O (20 mL), 10% H₂SO₄ (20 mL), and C₆H₆ (15 mL) were added to it and the C₆H₆ layer was separated after shaking the mixture. The aqueous layer was further extracted with C₆H₆ (15 mL×2), and the combined C₆H₆ extract was washed with H₂O, dried over Na₂SO₄ and evaporated down to dryness. The residue was chromatographed over silica gel column (35 g) eluted with CHCl₃-Me₂CO mixture. The fractions eluted with CHCl₃-Me₂CO (99:1) were further purified with a small silica gel column to give (-)-7 (126 mg, 42%) [α]_D²⁵=-18.9 (c 1.06, CHCl₃). The fractions eluted with CHCl₃-Me₂CO (95:5) were further purified with a small silica gel column to give (-)-5 (84 mg, 28%) with [α]_D²⁵=-19.2 (c 1.04, CHCl₃).
- (b) Reaction of (-)-4: The starting material, (-)-4 (300 mg, 0.87 mmol) dissolved in MeOH (10 mL) was reduced with NaBH₄ (343 mg, 8.7 mmol) and then oxidized with Al(Me₂CHO)₃ (246 mg, 1.0 mmol) as mentioned above. Work-up of the reaction mixture gave (+)-7 (130 mg, 43%) with $[\alpha]_D^{25}$ +18.7 (c 0.99, CHCl₃) and (+)-5 (83 mg, 28%) with $[\alpha]_D^{25}$ +19.1 (c 1.02, CHCl₃) after column chromatographic separation and purification.

Acknowledgements

The authors are grateful to NSC, R.O.C. for financial support of this research under Grants NSC 84-2331-B002-256 and NSC 85-2331-B002-245.

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- 6. Crystal data of (+)-8: monoclinic p21; a=5.6743 (II), b=14.254 (6), c=13.857 (6) Å, Z=2. Intensity data were collected on a CAD-4 diffractometer with θ/2θ scan mode, using monochromated MoKα radiation. Data were measured with 2θ range of 30–44°. A total of 2534 reflections were collected. Among them, 1489 were considered to be observed (>2.05 δ (I)). Final agreement indices are RF=0.059, RW=0.062, GoF=1.23, based on anisotropic refinement of all non-hydrogen atoms.
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(Received in Japan 16 June 1997; accepted 11 August 1997)